

Individual Variations in *Phlebotomus papatasi* Collected from Different Localities in Saudi Arabia

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ABSTRACT

A method of typing *Phlebotomus* taxa in three areas in Saudi Arabia (Riyadh, Madinah and Asir) using morphological characteristics revealed that the phlebotomine species is *Phlebotomus papatasi*. This identification was confirmed by establishing a polymerase chain reaction (PCR) and direct partial sequences of 18S ribosomal RNA (rRNA) gene using specific designed Primers SandF1:5'-AGGCTCATTTCAGTCGCTTTC-3' and SandR1:5'-TGCAAGCTTATGACTCACACTT-3'. Morphological individual variations were also observed among some collected specimens. Nevertheless, genetic analysis confirmed that these specimens were also *P. papatasi*. PCR-amplified amplicons using ChromasPro. MEGA 5 program and neighbor joining (NJ) methods revealed several direct sequences for *P. papatasi* that were submitted in GenBank under the accession number JQ929125. In conclusion, the obtained results establish a powerful tool for the molecular taxonomy of *Phlebotomus* spp. in endemic areas to plan appropriate epidemiological surveillance programs that could be used to detect natural infections of sand fly vectors with pathogens.

Keywords: Phlebotomine sand flies, PCR, 18S ribosomal RNA gene, direct sequences, morphological characters, molecular characters, taxonomy.

INTRODUCTION

Zoonotic cutaneous leishmaniasis is caused by the parasitic protozoan *Leishmania major*, which is transmitted by the phlebotomine sand fly *Phlebotomus papatasi* in North Africa and Middle East (Lane, 1993). Also *P. papatasi* has been implicated in the transmission of *L. arabica* in Saudi Arabia (Peters *et al.*, 1986; Kllick-Kendrick, 1990), and of arboviruses in many countries (Javadian *et al.*, 1977; Tesh *et al.*, 1977). It is postulated that successful establishment of the disease in an endemic area is the outcome of a close association between the *Leishmania* parasite and its natural sand fly vector (Hamarsheh, 2011). Thus, identification of both sand fly and *Leishmania* is of great importance for predicting expansion of the disease in endemic areas, and also it helps in designing new strategic programs that limit spreading of such serious vectors (Kato *et al.*, 2007; Fujita *et al.*, 2012).

Sand fly identification based on morphological characters includes terminal genitalia of males and internal structures of females, such as spermatheca and cibarium and also pharynx in the head region (Lango, 2005; Singh and Philips-Singh, 2010). On the other hand, the development of alternative molecular data has been recently introduced as tools for the identification of sand flies. Among these techniques are the ribosomal RNA (rRNA) gene architecture and the highly conserved sequences of certain domains of the gene. These rRNA gene sequences have been used to reevaluate higher level relationships within the subfamily Phlebotominae and within the genera *Phlebotomus* and *Sergentomyia* (Aransay, 2000). The rRNA gene is a multicopy gene of tandem repeated transcription units. Each transcription unit

includes regions that after processing produce three of the major rRNA subunits; those in insects include 18S, 5.8S, and 28S subunits) (Burton, 2008).

Therefore, the present study aims at identifying the phlebotomine sand flies collected from three localities in Saudi Arabia (Riyadh, Madinah, and Asir) using morphological and molecular characters (direct sequence of 18S rRNA gene subunit).

MATERIALS AND METHODS

Sand Flies Collection and Identification

Sand fly adults of both female and male sexes ($N = 250$) were collected, using sticky and light traps, during the year 2012 from three field localities (Riyadh, Al-Madinah, and Asir) (Table 1 and Fig.1). These localities were chosen based on the fact that they are the endemic areas of cutaneous leishmaniasis in Saudi Arabia (Lewis and Buttiker, 1980; Killick-Kendrick *et al.*, 1985). Populations were identified morphologically based on the previous morphological studies (Lewis and Buttiker, 1980; Killick-Kendrick *et al.*, 1985; El-Sibae and Eesa, 1993; Al-Dawood *et al.*, 2004). Immediately after sand fly collection, specimens were preserved in 70% ethanol. Each fly was dissected into many parts; the first part consists of the head and terminalia which was used for morphological identification according to the morphological taxonomic keys of (Lango, 2005; Carvalho and Jose, 2006; Alves *et al.*, 2008; Singh and Philips-Singh, 2010). While the other body parts of each fly were stored at -80°C for the molecular identification.



Fig. 1: Saudi Arabia political map showing Riyadh, Madinah and Asir regions (www.mapsofworld.com) (updated on, 16th May, 2012).

Table 1: *Phlebotomus papasi* prevalence in some Saudi Arabian localities during the year 2012.

Locality	Males	Females
Central region of Riyadh and its surrounding areas	30	2
Western region of Madinah and its surrounding areas	25	10
Southern region of Asir and its surrounding areas	58	7
Total	113	19

DNA Extraction

Individual ethanol-fixed specimens were homogenized and lysed in a DNA extraction kit (DNeasy tissue kit, Qiagen, California, USA) following manufactures instructions. Then, 5 μL portions of the DNA extracts were subjected to the polymerase chain reactions (PCRs) amplification.

PCR-Gene Amplification and Sequencing

The primers SandF1:5'-AGGCTCATTTCAGTCGCTTTC-3' and SandR1:5'-TGCAAGCTTATGACTCACACTT-3' was designed to bind within highly conserved regions. A region of 540 bp was amplified, of which a central 200 bp portion was predicted to be diagnostic. PCRs were performed in a final volume of 50 μ L. The products were resolved on a 2% (w/v) agarose gel and visualized under UV transilluminator and submitted directly for sequencing with the same F1 and R1 primers using the sequencing kit, Big Dye terminator V3.1 cycle (cycle sequencing kit, Applied Biosystems, Foster city, California, USA). Sequences were analyzed using ABI 3700 DNA analyzer (Applied Biosystems, Foster city, California, USA) (Aransay *et al.*, 1999). The consensus sequences for the Saudi Arabian *P. papatasi* have been deposited in GenBank, with accession number JQ929125.

RESULTS

Morphological Characteristics

Of the collected sand fly specimens, 132 specimens (52.8%) were identified to belong to the *Phlebotomus papatasi* (Table 1), where all individuals are characterized by having hairs with round sockets (Fig. 2). *P. papatasi* was the most abundant species and it was collected from the three studying areas. Males belonging to this species have long style with five spines, three terminal spines, and two lateral. The distance between the lateral spines is less than that between the lateral and terminal ones (Fig. 3a). Aedagus with round tip where the sperm duct pass through it (Fig. 3a). Paramere is trilobed, lateral lobes are shorter than the coxite and each of them bears two terminal spines (Fig. 3a). Female pharynx with narrow anterior and wide posterior ends, and armed with longitudinal ridges and several rows of teeth in the posterior part (Fig. 3b). Females are also characterized by having unarmed cibarium (Fig. 3c). Spermatheca is divided into ten segments. The terminal segment is the largest and the rest of the preceding ones become smaller that terminate with long segmented spermathecal duct (Fig. 3d).

However, 15 male specimens showed individual variations; (eight) specimens collected from (Al-Madinah) showed styles, each with six spines; three terminal, and three lateral spines (Fig. 4a). However, (seven) specimens collected from (Asir) exhibited two lateral lobes with different numbers of terminal spines. The formula of these spines for the two lobes was 2 + 3 (Fig. 4b), 3 + 3 (Fig. 4c), and 2 + 1 (Fig. 4d), respectively.

Molecular Characteristics

Analysis of individual sequences belonging to *P. papatasi* species showed little differences in the 18S rRNA subunit among the randomly chosen twelve individual sequences and found in match with the GenBank reference AJ244414.1 in the region between nucleotides 1 to 20 only (Fig. 5). PCR-amplified amplicons using ChromasPro. MEGA 5 program and neighbor-joining (NJ) methods revealed direct sequences for *P. papatasi* that are submitted in GenBank under the accession number JQ929125.

DISCUSSION

The morphological features of sand fly adults such as the chaetotaxy on the style and lateral lobes of terminal genitalia in males, as well as the internal features such as the spermatheca, cibarium, and pharynx in females confirmed that the identified

phlebotomine sand fly species is *P. papatasi*. These characteristics are in agreement with the previous morphological identification of *P. papatasi* (Lewis and Buttiker, 1980; Killick-Kendrick *et al.*, 1985; El-Sibae and Eesa, 1993; Al-Dawood *et al.*, 2004). Nevertheless, damage caused by improper storage of specimens and mounting may make the process of identification difficult or can cause misidentification.

Therefore, molecular characteristics markers have been explored in the present investigation for the development of more accurate techniques to confirm the morphologically identified sand flies collected from different localities in Saudi Arabia. The results of genetic analysis in the present study confirmed the accepted classification based on morphological characteristics even with the presence of morphological individual variations.

However, discrepancies existing between the two tools of classification, suggesting the necessity for careful reconsideration of the sand fly taxonomy. Therefore, genotyping method was shown to be accurate and easy-to-use for the identification of sand fly species, requiring less expertise and less risk of different interpretations among researchers than the conventional morphology-based classification. It is important to note that this DNA-based technique does not require special storage conditions for the specimens, and different methods of preservation such as drying and the use of 70% ethanol or liquid nitrogen, which may affect the quality of the results. In addition, damage of samples, which should affect the morphologic classification in many cases, does not affect the genotyping analysis (Kato *et al.*, 2007; Baroso *et al.*, 2007; Terayama *et al.*, 2008).

In conclusion, the obtained results establish a powerful tool for the molecular taxonomy of *Phlebotomus* spp. in endemic areas to plan appropriate epidemiological surveillance programs that could be used to detect natural infections of sand fly vectors with pathogens.

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REFERENCES

- Al-Dawood, A. S.; Alahmed, A. M. and Kheir, S. M. (2004). Population dynamics of sandflies (Diptera: Psychodidae) in Hanifah valley, Riyadh, Saudi Arabia. *Pak. J. Biol. Sci.*, 7: 464- 467.
- Alves, V. R.; Rui, A. F. and Barrett, T. (2008). *Lutzomyia maruaga* (Diptera: Psychodidae), a new bat-cave sand fly from Amazonas, Brazil. *Memorias do Instituto Oswaldo Cruz*, 103: 251-253.
- Aransay, A. M.; Scoulica, E. T. and Selentis, Y. (2000). Phylogenetic relationships of phlebotomine sandflies inferred from small subunit nuclear ribosomal DNA, *Insect Molec. Biol.*, 9: 157-168.
- Aransay, A. M.; Scoulica, E. and Chaniotis, B. (1999). Typing of sandflies from greece and cyprus by DNA polymorphism of 18s rRNA gene. *Insect Molec. Biol.*, 8:179-184.
- Barroso, P. A.; Marco, J. D. and Kato, H. (2007). The identification of sandfly species, from an area of Argentina with endemic *leishmaniasis*, by the PCR-

- based analysis of the gene coding for 18s ribosomal RNA. *Ann. Tropic. Medic. and Parasitol.* , 101, 247-253.
- Burton, E. T. (2008). *Molecular biology genes to proteins*. 3rd edition , Jones and Bartlett Publishers, Inc., London, pp. 670.
- Carvalho, G. M. and Jose, D. A. F. (2006). Taxonomic revision of phlebotomine sand fly species in the series devise and anamnesis of the subgenus: *Psychodopygus mangabeira*, 1941 (Diptera: Psychodidae: Phlebotominae). *Memorias do Instituto Oswaldo Cruz*, 101: 129-136.
- EL-Sibae, M.M. and Eesa, N. M. (1993). A study on *Phlebotomus* species, the vector of leishmaniasis in Gassim, Saudi arabia. *J. Egypt. Societ. Parasitol.*, 23: 231-238.
- Fujita, M.; Kato, H. and Hashiguchi, W. (2012). Genotyping of sand fly species in Peruvian Andes where leishmaniasis is endemic. *Acta Tropica* 121: 93-98.
- Hamarasheh, O. (2011). Distribution of *Leishmania major* zymodemes to populations of *Phlebotomus papatasi*. a review. *Parasit. Vect.*, 4: 9.
- Javadian, E.; Tesh, R.; Saidi, S. and Nadim, A. (1977). Studies on the epidemiology of sand fly fever in Iran. III. Host-feeding patterns of *Phlebotomus papatasi* in an endemic area of the disease. *Am. J. Trop. Med. Hyg.*, 26: 294-298.
- Kato, H.; Uezato, H. and Gomez, E. H. (2007). Establishment of a mass screening method of sand fly vectors for *Leishmania* infection by molecular biological methods. *Am. J. Trop. Med. Hyg.*, 77: 324-329.
- Killick-Kendrick, R. (1990). Phlebotomine vectors of the leishmaniasis: a review. *Med. Vet. Entomol.*, 4: 1-24.
- Killick-Kendrick, R.; Leaney, A. J. and Bray, R. S. (1985). Zoonotic cutaneous leishmaniasis in Saudi Arabia: the incrimination of *Phlebotomus papatasi* as the vector in the Al-Hassa Oasis. *Trans. R. Soc. Trop. Med. Hyg.*, 79: 252-255.
- Lane, R.P. (1993). Sand flies (Phlebotominae). In *Medical Insects and Arachnids*. Edited by: Lane RP, Crosskey RW. London: Chapman and Hall; 1993:78-119.
- Lango, I. K. (2005). Structure and function of the spermathecal complex in the phlebotomine sandfly *Phlebotomus papatasi* scopoli (Diptera: Psychodidae): I. Ultrastructure and histology. *J. Biosci.*, 30:711-731.
- Lewis, D. J. and Buttiker, W. (1980). Fauna of Saudi Arabia. V2. Insects of Saudi Arabia Diptera: Fam. Psychodidae, Sub Fam. Phlebotomine, pp. 252-285.
- Peters W.; Elbihari, S. and Evans, D. A. (1986). *Leishmania* infecting man and wild animals in Saudi Arabia. 2. *Leishmania arabica* n. sp. *Trans. R. Soc. Trop. Med. Hyg.*, 80: 497-502.
- Singh, N. S. and Philips-Singh, D. A. (2010). Study on genitalia of phlebotominae sand flies (Phlebotomidae: Diptera) in Northern India: a new tool for detection of species . *J. Entomol.*, 7: 235-239.
- Terayama, Y. Kato, H. and Gomez, E. A. (2008). Molecular typing of sand fly species (Diptera, Psychodidae, Phlebotominae) from areas endemic for leishmaniasis in Ecuador by PCR-RFLP of 18S ribosomal RNA gene , *J. vet. Med. Sci.*, 70, 907-913,2008.
- Tesh, R.; Saidi, S.; Javadian, E. and Nadim, A. (1977). Studies on the epidemiology of sand fly fever in Iran. I. Virus isolates obtained from *Phlebotomus*. *Am. J. Trop. Med. Hyg.*, 26: 282-287.

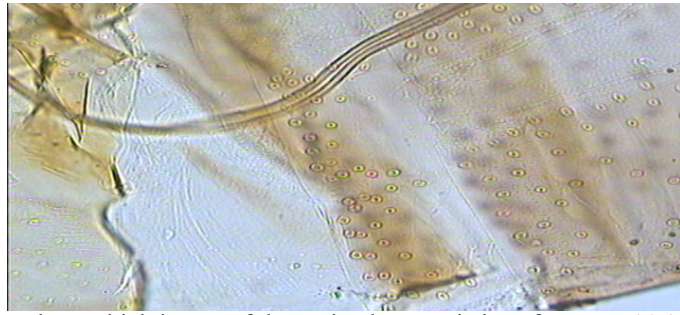


Fig. 2: Round hair sockets which is one of the main characteristics of genus *Phlebotomus* (40 ×).

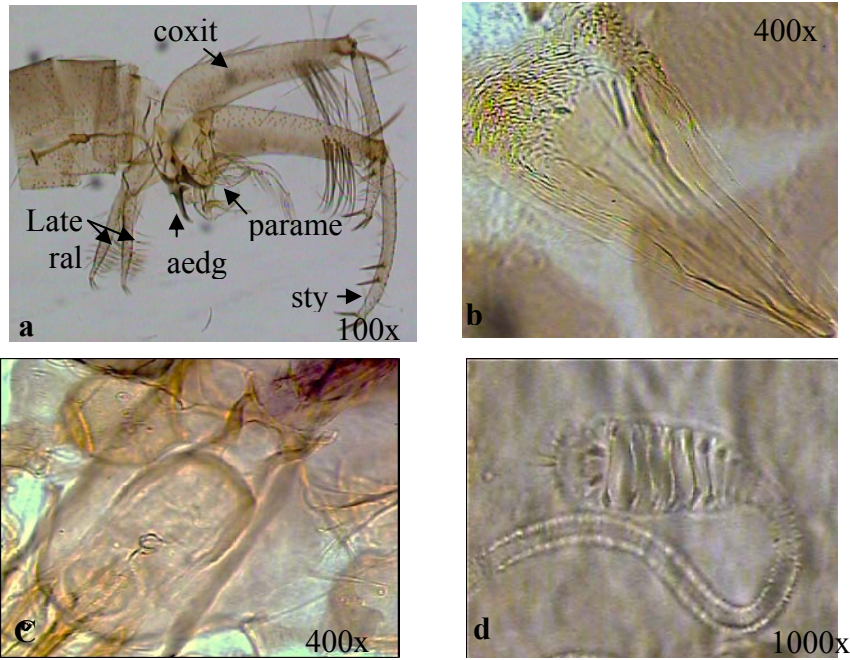


Fig. 3: Photomicrograph showing *P. papatasi* adult: (a) male terminalia; (b) female pharynx; (c) female cibarium; (d) female spermatheca.

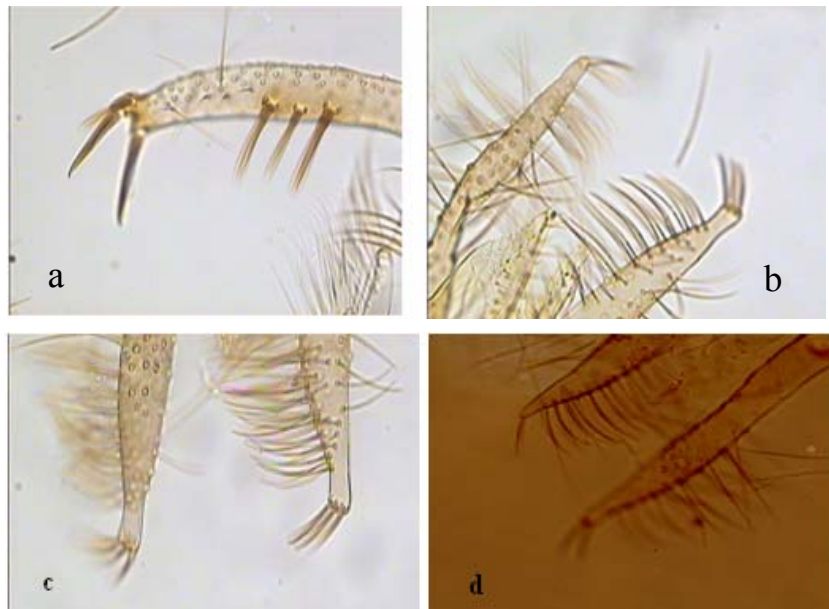


Fig. 4: Individual variations in *P. papatasi* males: (a) style with 6 spines, 3 terminal and 3 lateral (40 ×); (b) lateral lobes, one with two terminal spines and the other with three terminal spines (40 ×); (c) lateral lobes, each with three terminal spines (40 ×); (d) lateral lobes, one with two lateral spines and the other with only one lateral spine (40 ×).

10 20 30 40 50 60
|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|

AJ244414.1 **AGCTTTTGGGCTCCGGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGAA**
 P.pap1 **.ATCC.G.CA...A-C...G.....**
 P.pap2 **CAT...G.....--...G.....**
 P.pap3 **--A.....GA.TC...G.....-**
 P.pap4 **.AAG..G.....--...G.....**
 P.pap5 **..G.....--...G.....**
 P.pap6 **G.....--...G.....**
 P.pap7 **CA.A.....--...G.....**
 P.pap8 **-AACG.G.....--...G.....**
 P.pap9 **-.AC.....-...G.....-**
 P.pap10 **--GAC.....--...G.....**
 P.pap11 **-.ACG.....--...G.....-**
 P.pap12 **-CACG.....--...G.....**

 70 80 90 100 110 120
|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|

AJ244414.1 **GGGCACCACCAGGAGTGGAGCCTGCGGCTTAATTTGACTCAACACGGGAAAACCTTACCAG**
 P.pap1
 P.pap2
 P.pap3
 P.pap4
 P.pap5
 P.pap6
 P.pap7
 P.pap8
 P.pap9
 P.pap10
 P.pap11
 P.pap12

 130 140 150 160 170 180
|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|

AJ244414.1 **GTCCGAACATAAATGAGTAAGACAGATTGATAGCTCTTCTCGAATCTATGGGTGGTGGT**
 P.pap1
 P.pap2
 P.pap3
 P.pap4
 P.pap5
 P.pap6
 P.pap7
 P.pap8
 P.pap9
 P.pap10
 P.pap11
 P.pap12

 190 200 210 220 230 240
|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|.....|

AJ244414.1 **GCATGGCCGTTCTTAGTTCGTGGAGTGATTTGTCTGGTTAATTCGATAACGAACGAGAC**
 P.pap1
 P.pap2
 P.pap3
 P.pap4
 P.pap5
 P.pap6
 P.pap7
 P.pap8
 P.pap9
 P.pap10
 P.pap11

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P.pap12 .....
          250      260      270      280      290      300
          ....|....|....|....|....|....|....|....|....|....|....|....|
AJ244414.1 TCAAACTTTTTTAAATAGATGCTTGCAGGACTATGACGCTGAAGCCATGTGTATGTTGGT
P.pap1 .....
P.pap2 .....
P.pap3 .....
P.pap4 .....
P.pap5 .....
P.pap6 .....
P.pap7 .....
P.pap8 .....
P.pap9 .....
P.pap10 .....
P.pap11 .....
P.pap12 .....
          310      320      330      340      350      360
          ....|....|....|....|....|....|....|....|....|....|....|
AJ244414.1 TGCTCTGTTTTGCTTTCGGGTTAAATGGAGTGGTTTGATGTATATATGGTGGAGTCATA
P.pap1 .....
P.pap2 .....
P.pap3 .....
P.pap4 .....
P.pap5 .....
P.pap6 .....
P.pap7 .....
P.pap8 .....
P.pap9 .....
P.pap10 .....
P.pap11 .....
P.pap12 .....
          370      380      390      400      410      420
          ....|....|....|....|....|....|....|....|....|....|
AJ244414.1 CCTGTTTGGTTTGTCTTAAGAGGACACTTTGCTTCTTAAATGGACAAAATGCGTCTAGCA
P.pap1 .....
P.pap2 .....
P.pap3 .....
P.pap4 .....
P.pap5 .....
P.pap6 .....
P.pap7 .....
P.pap8 .....
P.pap9 .....
P.pap10 .....
P.pap11 .....
P.pap12 .....
          430      440      450      460      470      480
          ....|....|....|....|....|....|....|....|....|....|
AJ244414.1 TTAATGAGATTGAGCAATAACAGGTCTGTGATGCCCTTAGATGTCCCTGGGCTGCACGCGC
P.pap1 .....
P.pap2 .....
P.pap3 .....
P.pap4 .....
P.pap5 .....
P.pap6 .....
P.pap7 .....
P.pap8 .....
P.pap9 .....
P.pap10 .....
          .....G.....S.C.

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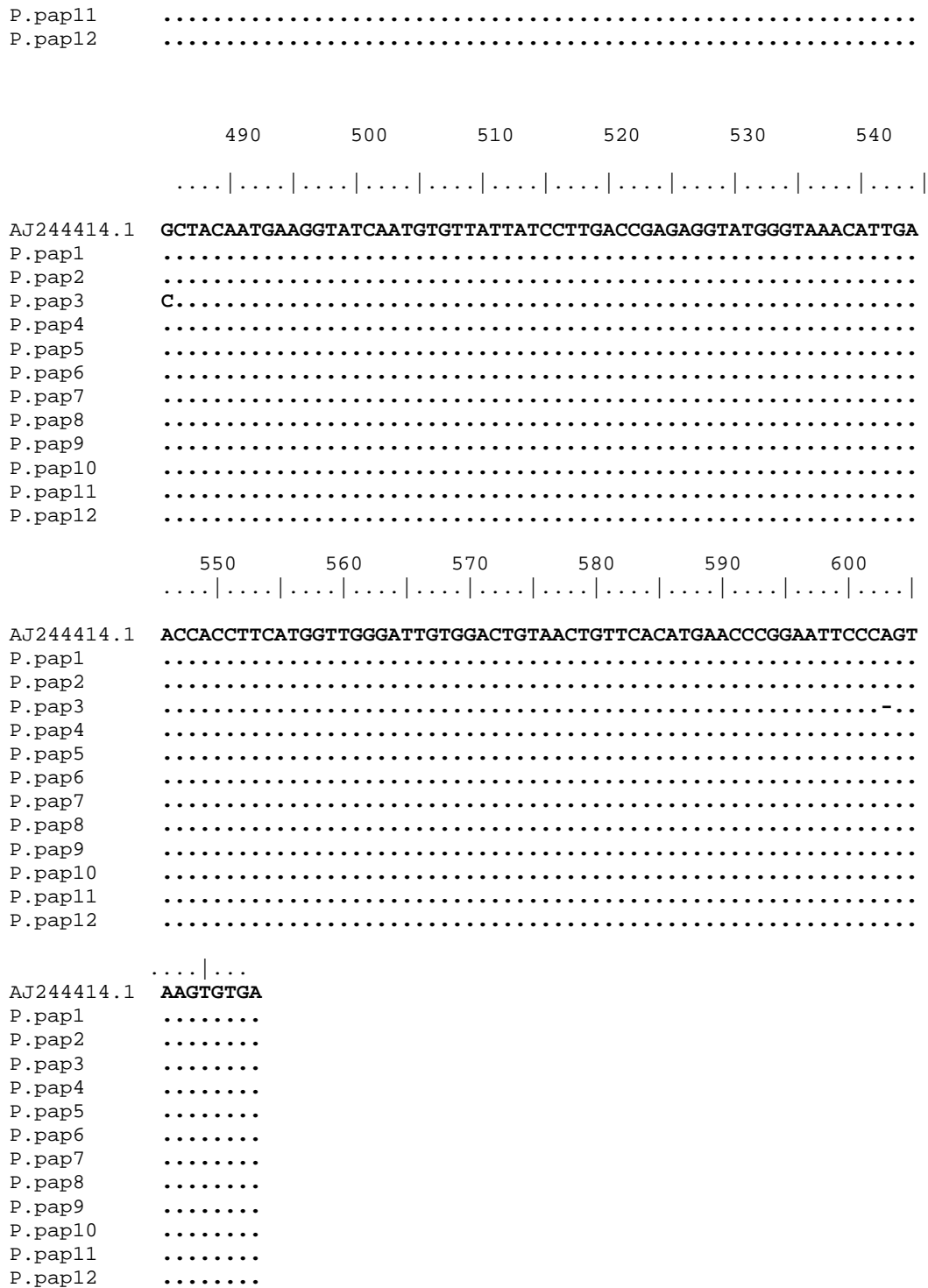



Fig. 5: 18S rRNA sequences alignment of *P. papatasi* individuals. Identities are denoted by dots and gaps by dashes. AJ244414.1 is a reference sequence.

ARABIC SUMMARY

الاختلافات الفردية في ذبابة الرمل التي *Phlebotomus papatasi* تم جمعها من مواقع مختلفة في المملكة العربية السعودية

ريم عطاالله العجمي

جامعة الملك سعود - كلية العلوم - قسم علم الحيوان - الرياض - أقسام العلوم والدراسات الطبية الملز

أوضحت طريقة تقسيم أنواع جنس *Phlebotomus* المنتشرة في ثلاث مناطق بالمملكة العربية السعودية ممثلة بالرياض والمدينة وعسير باستخدام الخصائص المورفولوجية أن نوع *Phlebotomus papatasi* هو أكثر الأنواع إنتشاراً.

وقد أكد ذلك التصنيف باستخدام تفاعل البلمرة المتسلسل المباشر (PCR) لسلاسل جزئية من الجين المشفر للحمض النووي الرايبوزي الريبوزومي 18S rRNA وذلك من خلال إستخدام بواقي مصممة متخصصة (S and F1:5'-AGGCTCATTTCAGTCGCTTTC-3' and S and R1:5'-TGCAAGCTTATGACTCACACTT-3'). هذا وقد لوحظ بعض الاختلافات الشكلية (المورفولوجية) الفردية بين بعض العينات التي تم تجميعها.

ومع ذلك أكد التحليل الجيني أن تلك العينات تنتمي الى نوع *Phlebotomus papatasi*. وقد كشفت الطرق التصنيفية الاحصائية والتقنية الحيوية المستخدمة [ChromasPro. MEGA 5 program and Neighbor-Joining (NJ)] تتابع جيني تسلسلي للجين المشفر للحمض النووي الرايبوزي الريبوزومي 18S rRNA والذي تم تسجيله في الجين بنك برقم JQ929125.

وعليه فإن النتائج التي تم الحصول عليها تعد نواة لأداة قوية للتصنيف الجزيئي لنوع *Phlebotomus* تجدد الاهتمام فيما يتعلق بتأثير إدراج هذه الأنواع في المناطق الموبوءة على خطة المراقبة الوبائية والبرامج المناسبة التي يمكن استخدامها للكشف عن الإصابة بذباب الرمل الناقل للجراثيم المسببة للأمراض الوبائية المختلفة.